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Description	

**A globotriaosylceramide (Gb₃Cer) mimic peptide isolated
from phage display library expressed
strong neutralization to Shiga toxins**

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Abstract

A Gb₃-trisaccharide mimic peptide was selected with biopanning from a phage display library against anti-Gb₃ antibody to neutralize Shiga toxins (Stxs). Biopanning was carried out on a microplate immobilized with a Fab fragment of anti-Gb₃ antibody and a subtraction procedure screening was applied to enhance specificity. The selected phage clones showed strong affinity to anti-Gb₃ antibody and to Stxs. Among these clones, a 9-mer sequence **WHWTWLSEY** was determined as the strongest Gb₃ mimic peptide and chemically synthesized. The peptide bound strongly to Stx-1 and Stx-2, though the binding was inhibited with Gb₃Cer. Surface plasmon resonance (SPR) and fluorescent spectroscopy determined that the affinity of the peptide to both Stxs was strong. Neutralization activity was confirmed by *in vitro* assay with HeLa cells. The Gb₃ mimic peptide potentially has great promise for use against Stxs.

Keywords: Carbohydrate mimic peptide, Gb₃, Phage display library, Shiga toxin

Abbreviations

ABTS, 2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid); BOP, benzotriazole-1-yloxy-tris(dimethylamino)phosphonium hexafluorophosphate; BSA, bovine serum albumin; *B. simplicifolia*, *Bandeiraea simplicifolia*; CBB, Coomassie Brilliant Blue; Cer, ceramide; Con A, concanavalin A; DMA, *N, N*-dimethylacetamide; DMEM, Dulbecco's modified Eagle's medium; ELISA, enzyme-linked immunosorbent assay; FITC, fluorescein isothiocyanate; Fmoc, fluorenylmethoxycarbonyl; Gb₃, globotriaosylceramide Gal(α1-4)Gal(β1-4)GlcβCer; HBS-EP, HEPES-buffered saline with EDTA and polysorbate added; HEPES, 4-(2-hydroxyethyl)-piperazine-1-ethanesulfonic acid; HOBt, 1-hydroxybenzotriazole; HPLC, high-performance liquid chromatography; HRP, horseradish peroxidase; Ig, immunoglobulin; WST-8, 2-(2-methoxy-5-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2*H* tetrazolium monosodium salt; MALDI, matrix-assisted laser deposition ionization; RCA₁₂₀, *Ricinus communis agglutinin* 120; RMS, root mean square; SPR, surface plasmon resonance; Stx, Shiga toxin; TBS, Tris buffered saline; TOF, time of flight.

1. Introduction

Because bioactive saccharides play important roles in the molecular recognition of living processes such as cell adhesion, tumor metastasis and infection, they have attracted much interest for their medicinal and pharmaceutical applications [1,2]. However, there are problems in utilizing the carbohydrates as drugs and epitopes, such as the difficulty obtaining sufficient quantities of pure oligosaccharide from either natural sources or synthetic procedures, and the weak carbohydrate-protein interaction.

The utilization of carbohydrate-mimetic compounds is a promising method for overcoming these difficulties [3-5]. Among the mimetic compounds, carbohydrate-mimic peptides have come in for much attention [6-20]. Carbohydrate-mimic peptides often show greater affinity for antibodies, enzymes and lectins than native carbohydrates. Carbohydrate mimic peptides can elicit an immune response that can be utilized for vaccine development, for example against diseases as cancers. In addition, peptides can be cloned within a short time from a phage display peptide library and can be synthesized by a simple solid-phase strategy that is superior to the more tedious process of saccharide synthesis. Carbohydrate-mimic peptides have been analyzed in terms of both structure and function, and investigations of them have increased remarkably.

Here, we report the production of Gb₃ mimic peptides by phage display library to

develop a drug to neutralize Shiga toxins (Stxs: Stx-1 and Stx-2). Stxs produced by *Eschericia coli* O157:H7 cause serious clinical complications [21]. Stxs are multisubunit proteins consisting of one toxic A-subunit and five carbohydrate-binding B-subunits [22]. The functional ligand for Stxs is Gb₃Cer glycolipid (α -D-Gal(1-4) β -D-Gal(1-4) β -D-Glc-Ceramide), which mediates the internalization of Stxs into host cells [23]. There have been extensive studies on the molecular design of artificial Stxs ligands using globotriose (a trisaccharide unit of Gb₃Cer), including our own reports on globotriose-carrying polymers with strong neutralization activities [24]. Since the interactions between carbohydrate mimic peptides and lectins are usually stronger than those between carbohydrates and lectins, the Gb₃ mimic peptides have the potential to be developed for new medicinal applications.

To select a Gb₃ mimic peptide, we adopted biopanning using a phage display to monoclonal anti-Gb₃ antibody, and then examined the Gb₃ mimic peptide with a subtraction procedure screening. We synthesized Gb₃ mimic peptides and investigated their interactions with Stxs by enzyme-linked immunosorbent assay (ELISA), surface plasmon resonance (SPR) and fluorescent spectroscopy. Gb₃ mimic peptides were also analyzed by computer simulations. Our detailed information expands the potential of carbohydrate mimic peptides for biomedical applications.

2. Materials and methods

2.1. Chemicals

The following substances were used as received: anti-Gb₃ antibody (rabbit, IgG2b) (Seikagaku, Tokyo, Japan); Shiga toxins (Stxs) and Gb₃Cer (Nacalai Tesque, Kyoto, Japan); concanavalin A (Con A) and *Ricinus communis* agglutinin 120 (RCA₁₂₀) (Honen Co. Ltd., Tokyo, Japan); *Bandeiraea simplicifolia*, papain agarose, iodoacetamide and Gly-HCl (Sigma-Aldrich, St. Louis, MO, USA); anti-M13 antibody and bovine serum albumin (BSA) (Amersham Bioscience, Piscataway, NJ, USA); 1-hydroxybenzotriazole hydrate (HOBt) and benzotriazole-1-yloxy-tris(dimethylamino)phosphonium hexafluorophosphate (BOP) (Kokusan Chemical, Tokyo, Japan); fluorescein-4-isothiocyanate (FITC-I) and 2-(2-methoxy-4-nitrophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2H tetrazolium monosodium salt (WST-8) (Dojindo, Kumamoto, Japan); and Fmoc-amino acids (Peptide Institute, Osaka, Japan).

2.2. Antibody purification [25]

The Fab fragment of the anti-Gb₃ antibody was prepared as follows. Anti-Gb₃ antibody

(50 μg), 1 M cysteine (6 μl), 20 mM EDTA (6 μl) and papain agarose (1.6 μl) were dissolved in acetate buffer (pH 5.5, 50 μl) in a tube and mixed with a tube rotator for 45 min. Papain was deactivated with 1 M iodoacetamide (4 μl) and the resultant solution was centrifuged twice at 15000 rpm at 4 °C for 10 min. The production of Fab fragment was analyzed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) using 10 % gels. Gels were stained with Coomassie Brilliant Blue (CBB). The supernatant was separated from precipitated papain. pH was adjusted to 8.0 with Tris buffer (pH 8.5). Protein A agarose was added to the solution and incubated at 4°C for 3 h to remove the Fc fragment. The solution was centrifuged at 15000 rpm at 4 °C for 10 min to obtain the Fab solution as supernatant. The concentration of Fab was determined by absorbance at 562 nm, comparing the absorbance of Fab of IgG2.

2.3. Biopanning with anti-Gb₃ antibody

Biopanning was carried out using a phage display library kit (New England Biolabs., Cambridge, MA, USA) based on a combinatorial library of random 12-mer peptides fused to a minor coat protein (pIII) of the filamentous coliphage M13. A 96-well plate was coated with 100 μg of anti-Gb₃ Fab at 4 °C overnight, treated with 200 μl of blocking solution (100 mM NaHCO₃, 5 mg/ml BSA, and 0.02 % NaN₃) at 4 °C for 2 h, and washed

six times with Tris buffered saline (TBS) containing 0.1 % Tween. The phage library (1.5×10^{11} pfu) was added to the plate and incubated at room temperature for 60 min. The plate was washed six times with TBS 0.5 % Tween, and the bound phages were eluted with 0.1 N Gly-HCl buffer (pH 2.2). The eluate was neutralized with 1 M Tris HCl buffer. The phage clones obtained were then amplified by infecting them into host bacterial cells (*E. coli* ER 2738).

2.4. Screening utilizing a phage display subtraction method [26]

Subtraction biopanning was independently carried out. A 96-well plate was coated with 100 μ g of IgG2 in 100 μ l of 0.1 M NaHCO₃ buffer (pH 8.6) at 4 °C overnight, treated with 200 μ l of blocking solution at 4 °C for 2 h, and washed six times with TBS 0.1 % Tween. The phage library (1.5×10^{11} pfu) was added to the plate and incubated for 60 min. The supernatant containing unbound phage clones was collected and purified, and the unbound phage was subject to biopanning with anti-Gb₃ antibody by the above-detailed method.

2.5. ELISA

Isolated phage clone binding to anti-Gb₃ antibody and Stxs was analyzed by ELISA. A

96-well plate was incubated with anti-Gb₃ antibody (70 µl of 70 µg/ml in 0.1 M NaHCO₃ (pH 8.6)) at 4 °C for 14 h, or with Stxs (70 µl of 10 µg/ml in PBS buffer) at 4 °C for 16 h. The antibody or Stx solution was removed, and the plate was incubated with 200 µl blocking buffer at 4 °C for 2 h. The plate was washed six times with TBS-0.5% Tween, and the phage clones (1.6 x 10⁹ virions for anti-Gb₃ antibody and 2.4 x 10⁷ virions for Stxs) were added to the plate. After incubation for 1 h, the plate was washed six times with TBS-0.5% Tween. Horseradish peroxidase (HRP) anti-M13 conjugate in blocking buffer was added and the plate was incubated for 30 min. The assay was visualized by addition of H₂O₂ and 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS) followed by quenching with sodium citrate for 1 h. Absorbance at 405 nm was recorded as a function of the diluted phage with a microplate reader. All results were averages of at least four different measurements.

An aqueous solution of Gb₃Cer was prepared by vortex mixing of Gb₃Cer (100 µg) with PBS buffer (50 ml) for overnight. Anti-Gb₃ antibody or Stxs were incubated with the aqueous solution of Gb₃Cer (0.5 µl of 2 µg/ml) for 1 h, followed by assay of the mixture of phage and Gb₃Cer according to the above-detailed method.

2.6. Peptide synthesis

Peptides were synthesized by a solid phase method using standard Fmoc chemistry on Wang resin. Coupling of amino acids was carried out using HOBt and BOP in DMA. The Fmoc group was removed in 20 % piperidine/DMF. FITC labeled peptide was synthesized by incubation of peptide (**AcWHWTWLSEYK**) with fluorescein-4-isothiocyanate (FITC) [27]. Peptides were purified by reversed phase HPLC (JASCO, Tokyo, Japan) with a gradient of acetonitrile/0.1 % TFA; purity as estimated by HPLC was >95 %. MALDI-TOF-MS: **WHWTWLSEY** 1330.7 [M+Na]⁺ (calculated 1330.4), **AcWHWTWLSEYK** 1478.1 [M+H]⁺ (calculated 1478.6). **AcWHWTWLSEYK-FITC** 1822.9 [M+H]⁺ (calculated 1822.7).

2.7. Peptide-protein interaction

Interactions of the Gb₃ mimic peptide with proteins were analyzed by SPR and fluorescence spectroscopy, with a Langmuir binding model assumed for both analyses. SPR was carried out with a BIAcore 2000 using CM5 chip (BIAcore, Uppsala, Sweden) on which a peptide having Lys at the C-terminal (**AcWHWTWLSEYK**) had been immobilized. Measurements were carried out in HBS-EP buffer (pH 7.4) at 4 °C at a flow rate of 20 µl/min. Proteins at various concentrations were injected to determine

their affinity with the peptides. The chip surface was regenerated to remove bound analyte by injection of 5 mM Tris-HCl (pH 9.0) for 1.5 min. Kinetic constants were calculated with BIAevaluation software.

The dissociation constants were also analyzed by fluorescence changes of FITC-labeled peptide (AcWHWTWLSEYK-FITC) (Ex. 490 nm, Em. 518 nm) with FP-6500 (JASCO, Tokyo, Japan). Proteins with various concentrations were added to the peptide solution in PBS buffer at 4 °C, and the interactions were analyzed by Scatchard plots [28].

2.8. Neutralization of Stxs with a Gb₃ mimic peptide [29]

Neutralization activity of the Gb₃ mimic peptide was examined *in vitro* using a HeLa cell cytotoxicity assay. HeLa cells were cultured in Dulbecco's modified Eagle's medium (DMEM) containing 10 % fetal bovine serum (FBS) at 37 °C for 24 h. An aliquot containing 5 x 10³ cells was transferred to each well of a 96-well plate, and DMEM was removed. A solution of Stx (5 pg) and the Gb₃ mimic peptide (0.001 – 100 µg) in 100 µl DMEM, preincubated at room temperature for 2 h, was added to each well. The cells were incubated in a 5 % CO₂ atmosphere at 37 °C for 72 h. Cell indicator WST-8 (10 µl) was added to the plate, and the cell survival rate was calculated by absorbance at 480

nm. Results were calculated from four different wells.

2.9. Computer simulation

Molecular simulations of the binding of a Gb₃ mimic peptide (**WHWTW**) to a Stx-1 B subunit were carried out using Amber 6 (University of California, San Francisco, CA, USA) on the basis of the model in the Protein Data Bank (PDB 1BOS) [30]. The binding was first calculated by random modeling with a genetic algorithm and then, optimized under an Amber 1994 force field. The water was appended in the Generalized-Born model. The potential energy of the structures was minimized with a root mean square (RMS) gradient of 0.01 kcal mol⁻¹ Å⁻¹ [31].

3. Results

3.1. Selection of anti-Gb₃ antibody binding phage clones.

We screened the peptides bound to anti-Gb₃ antibody. To efficiently select the Gb₃ mimic peptide, the antibody Fab fragment was liberated through hydrolysis of the hinge region with papain and purified. After three rounds of biopanning, 10 individually isolated bacterial clones were randomly selected, and the phage clones obtained from the host bacterium were subjected to DNA sequence analysis. The deduced peptide

sequences are summarized in Table 1a. The sequences of the 10 colonies were classified into seven clones. Clones **Gb3-1** from three colonies, **Gb3-3** and **Gb3-4** were found to carry a 6-mer consensus sequence **WXWTWTXL** (X=H or Q). **Gb3-1** and **Gb3-3** carried a similar 12-mer sequence **WHWTWLSXYPPP** (X=E or D). Biopanning to Stx-1 was also investigated but to no avail; a consensus sequence with Stx-1 affinity was not obtained.

3.2. Analyses of the selected clones by ELISA

The binding of the selected phage clones to anti-Gb₃ antibody was assayed by ELISA (Fig. 1a). Clones **Gb3-1**, **Gb3-3**, **Gb3-4** and **Gb3-7** were well bound to anti-Gb₃ antibody, which contrasted with the modest binding of **Gb3-5** and the low binding of **Gb3-2** and **Gb3-6**. ELISA was carried out in the presence of an excess amount of Gb₃Cer to confirm Gb₃ mimicry. Binding of **Gb3-1**, **Gb3-3** and **Gb3-7** was inhibited by Gb₃Cer, but that of **Gb3-4** was inhibited just a little.

The extent of phage clone binding to Stx-1 and Stx-2 was analyzed by ELISA (Fig. 1b). The phage clones having a consensus sequence of **WXWTWL** (X=H or Q) (**Gb3-1**, **Gb3-3** and **Gb3-4**) showed a lot of binding to both Stxs, and binding was inhibited with an excess amount of Gb₃Cer. **Gb3-1** had the greatest amount of binding. **Gb3-5** had a high

amount of binding to Stx-1, but this was modestly inhibited by Gb₃Cer. These phage clones were strongly bound to both Stxs. **Gb3-7** carrying a different sequence also showed Gb₃ mimicry.

3.3. Screening using a subtraction method

Using the above conventional biopanning procedure, selected phage clones were probably bound not only to the variable domain but also to the constant domain of the Fab fragment. Subtraction screening using IgG2 as a negative control was carried out to enhance the binding specificity to the Fab fragment variable domain. After four rounds of biopanning six different sequences were obtained from 10 colonies (Table 1b) and subjected to ELISA. **Gb3-S-1** was the only clone that showed a great extent of binding to anti-Gb₃ antibody. The **Gb3-S-1** clone obtained from three colonies was found to carry the 12-mer **WHWTWLSEYPPP** sequence identical to that of the **Gb3-1** clone in Table 1a. We believed that **Gb3-1** bound most selectively and strongly to the variable domain of anti-Gb₃ antibody.

3.4. Binding affinity of the Gb₃ mimic peptide

We have selected **WHWTWLSEYPPP** of the **Gb3-1** clone as the most important Gb₃

mimic sequence. Since C-terminal Pro-Pro-Pro was not concerned with interaction with Stxs and was a little difficult to synthesize, we synthesized 9-mer **WHWTWLSEY** as a Gb₃ mimic peptide based on **Gb3-1** clone. We also synthesized **AcWHWTWLSEYK** for the affinity measurements, in which Lys (K) at the C-terminal and an acetyl group at the N-terminal were designed to conjugate the peptide to the biochip of SPR and to FITC [32].

The affinity of the synthetic Gb₃ mimic peptide for Stxs was quantitatively evaluated by SPR (Fig. 2) [33]. The peptide had strong affinity for both Stx-1 and Stx-2. The binding response (RU) of Stx-1 was a little greater than that of Stx-2, but both were of the same order. The affinities for non-carbohydrate binding proteins such as trypsin and fibrinogen were very weak; each RU was less than 10 % of that for Stxs. The rate equilibrium constants of association and dissociation to Stxs were estimated based on assumptions of a heterogeneous binding model (Peptide + Stx → Peptide-Stx) (Table 2). The association rate constant k_a and dissociation rate constant k_d of the peptide for Stx-1 were larger and smaller, respectively than those for Stx-2. Consequently, the peptide showed stronger affinity to Stx-1 than to Stx-2.

Binding of the Gb₃ mimic peptide to Stxs was also analyzed by fluorescence spectroscopy with a FITC-labeled peptide. The dissociation constants for Stxs were

calculated from the changes in fluorescence as 3.6×10^{-7} and 5.2×10^{-7} (M). Although the binding affinities by fluorescence were a little weaker than that by SPR, fluorescence analysis also showed that the Gb₃ mimic peptide bound strongly to Stxs.

Unfortunately, we found that the Gb₃ mimic peptide also bound to other lectins. The dissociation constants of the peptide with *B. simplicifolia* (α -Gal recognition lectin), RCA₁₂₀ (β -Gal recognition lectin) and Con A (α -Glc/ α -Man recognition lectin) were 4.1×10^{-7} , 3.3×10^{-7} , 2.4×10^{-7} (M) in SPR and 5.6×10^{-5} , 3.8×10^{-5} and 3.1×10^{-5} (M) by fluorescence measurement.

3.5. *In vitro* neutralization of Stxs with the Gb₃ mimic peptide

The Gb₃ mimic peptide was evaluated for its activity neutralizing Stxs (Fig. 3). Cell survival rates against Stxs were increased by addition of the peptide. The addition of 10^3 μ g/ml peptide conferred sufficient protection on HeLa cells in the presence of Stxs. The protective effect on Stx-1 and Stx-2 was similar. The dilute peptide solution (10^{-2} – 1 μ g/ml) showed a stronger protective effect on Stx-2 than Stx-1, and the concentrated solution (1 – 10^3 μ g/ml) showed almost identical protection of Stx-1 and Stx-2. The protective effects of the peptide on HeLa cells were modest in spite of a small K_D value. The neutralization activity of the peptide in serum free medium was also investigated,

which exhibited better protective effects than that in the presence of serum.

3.6. Computer simulation of Stx-1 binding mode

Binding between Stx-1 and the peptide was simulated using the first penta-peptide (**WHWTW**) of the Gb₃ mimic peptide to analyze binding behavior and binding sites (Fig.

4). Binding was calculated using random modeling; we obtained 16 binding modes with low potential energies from 50 binding modes. The potential energy of the 16 binding modes was almost the same, within 12 kcal/mol, and the majority of the peptides (13 out of 16, ca 80 %) bound to site 2.

4. Discussion

We have been successful in the selection of a Gb₃-mimic peptide (**WHWTWLSEY**) based on biopanning for anti-Gb₃ antibody. As shown in Table 1, the selected consensus sequence was **WXWTWLS** (**X=H** or **Q**) (**Gb3-1**, **Gb3-3** and **Gb3-4**). Although peptides with the consensus sequence bound well to the anti-Gb₃ antibody, only those with a second His residue (**Gb3-1** and **Gb3-3**) were inhibited by Gb₃Cer, indicative of Gb₃ mimicry. Therefore the second His was considered to be essential. Besides, clones **Gb3-1** (**WHWTWLSEY**) and **Gb3-3** (**WHWTWLSDY**) had the same sequence except for the eighth residue, but **Gb3-1** bound more to Stxs than **Gb3-3**. The difference of only one

residue affected Gb₃ mimicry and Stxs binding. The Gb₃ mimic peptide (WHWTWLSEY) was the best ligand for Stxs, which was also confirmed by biopanning using the subtraction procedure. On the other hand, clones with the hydrophobic residues Gb3-5 and Gb3-7 also bound strongly to Stxs, suggesting the contribution of a hydrophobic interaction. Since these clones did not have the consensus sequence and were not selected by the subtraction procedure, Gb3-1 was more suitable for Gb₃ mimicry. Gb3-2 and Gb3-6 were selected by biopanning in spite of having a weak interaction with the anti-Gb₃ antibody and Stxs. These clones were selected due to the bias of the M13 phases [34].

The selected Gb₃ mimic peptide (WHWTWLSEY) has alternate hydrophobic (Trp) and hydrophilic residues (Thr, Ser, Glu and His). Trp can contribute to Stxs binding by a strong hydrophobic interaction [35] and π - π stacking. X-ray analysis showed that the α -anomeric surface of the Gal in Gb₃ was bound to Stx-1 through hydrophobic interaction with Leu29, Phe30 and Trp34 [30]. The Gb₃ mimic peptide amplifies the hydrophobic interaction due to its stronger hydrophobicity. The aromatic residues Phe30 and Trp34 in Stx-1 contribute to the π - π stacking with Trp in the Gb₃ mimic peptide, which was observed by computer simulation. Hydrophilic residues (Thr, Ser, Glu and His) of the Gb₃ mimic peptide can form hydrogen bonds with Stxs; this is

similar to the hydrogen bonding of Gal in Gb₃ with Thr21, Gly60, Asp17, Arg33 and Asp18 in Stx-1. His also stabilizes conformation for Stxs binding due to limited torsion angles [36].

The Gb₃ mimic peptide was predicted to bind to Stxs in the same way as Gb₃Cer, which was shown by binding inhibition with Gb₃Cer. It has been reported that Stx-1 has three types of Gb₃-binding sites (sites 1, 2 and 3) in a B-subunit, and that the binding affinity is in the order of site 2 > site 1 >> site 3 [30, 37-39]. Therefore the Gb₃ mimic peptide was expected to bind to site 2 in Stx-1. The interaction between Stx-1 and the peptide was analyzed by computer simulation using the first penta-peptide (WHWTW). Most results showed that the peptide bound to site 2 with many different binding modes, though the potential energy of the binding modes was almost the same. Site 2 is advantageous for entropic cooperativity due to its binding modes [40, 41].

A striking feature of the Gb₃ mimic peptide was its strong binding to both Stx-1 and Stx-2, while Gb₃Cer bound strongly to Stx-1 but only modestly to Stx-2 [25, 31, 33, 34]. The conformation of site 2 in Stx-2 differs from that in Stx-1, resulting in the strong binding of the Gb₃ mimic peptide to Stx-2 and a weak binding for Gb₃Cer. It has been reported that Stx-2 is more toxic than Stx-1 and that Gb₃ binds modestly to Stx-2 compared with Stx-1. Therefore the strong interaction of the peptide with Stx-2 is a

useful feature of Gb₃ mimic peptides.

The Gb₃ mimic peptide bound not only to Stxs, but also to other lectins. Considering the Gb₃ structure of Gal(α 1-4)Gal(β 1-4)Glc β Cer, the peptide binding to *B. simplicifolia* and RCA₁₂₀ was adequate based on the property of Gal recognition of these lectins. However, the Gb₃ mimic peptide also bound to Con A in spite of the α -Glc/ α -Man recognition property, suggesting indiscriminate binding. The Gb₃ mimic peptide was rich in hydrophobic residues, which induced the non-specific binding to α -mannose recognition lectin. Compared with Gb₃Cer, the Gb₃ mimic peptide exhibited similar or better neutralization activity to Stxs, but Gb₃Cer had better recognition specificity.

The protective effects of the peptide on Stxs were not strong in spite of the small K_D values; the unstable peptide conformation for Stxs binding is one possible reason for this. Since the synthetic 9-mer peptide (WHWTWLSEY) freely changes steric configurations in an aqueous solution, the frequency of forming a carbohydrate mimic structure seems to be very low, and so weakens binding affinity. But when peptides were immobilized on an SPR chip, the frequency of the mimicking Gb₃ structure seemed to increase; in fact, the dissociation constants obtained by fluorescence were larger than those obtained by SPR, indicating the weaker binding of the isolated peptide to Stxs. That confirms that, the microenvironment surrounding the peptide is important for the

peptide conformation for Stxs binding. Another possible reason is the non-specific binding of the peptide to other proteins. The protective effects of the peptide in serum free medium was better than that in the presence of serum, which suggests the peptide bound to other proteins in the serum non-specifically to reduce the protective effects. A multivalent compound or a peptide with a rigid conformation may overcome these defects.

5. Conclusion

A Gb₃ mimic peptide (WHWTWLSEY), based on the Gb3-1 clone selected from a phage display, was successfully prepared as a novel neutralizer for Stxs. A Gb3-1 clone was found to bind well to anti-Gb₃ antibody and Shiga toxins by ELISA. The affinity of the Gb₃ mimic peptide for Stx-2 was of the same order as that for Stx-1. The peptide neutralized Stxs modestly in HeLa cells *in vitro*. Investigation of efficient Stxs neutralization by a multivalent compound and the antibiotic peptide for *E. coli* O-157:H7 is currently under way.

Due to its great efficacy, the Gb₃ mimic peptide is a useful bioactive substance. We expect that the Gb₃ mimic peptide can efficiently express not only neutralizing activity but also other carbohydrate biological activities such as cell adhesion, cell growth and

immune response.

Supplementary Data

The result of ELISA in subtraction biopanning, the neutralization of Stxs in the serum free medium and the results of computer simulation are available.

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Figure Legends

Fig. 1 Binding assay of selected phage clones by ELISA (a) to anti-Gb₃ antibody in the absence and presence of Gb₃Cer (0.4 μg/ml in 200 μl PBS buffer) with 1.6 x 10⁹ virions of phage clones, and (b) to Stxs in the absence and presence of Gb₃Cer (20 μg/ml in 200 μl PBS buffer) with 2.4 x 10⁷ virions of phage clones. Experiments were repeated at least four times.

Fig. 2 SPR trace for binding of a Gb₃ mimic peptide (**AcWHWTWLSEYK**) to Stx-1, Stx-2, fibrinogen, and trypsin at 25°C at a protein concentration of 50 μg/ml.

Fig. 3 Neutralization of Stxs with a Gb₃ mimic peptide (**WHWTWLSEY**). Survival rates of HeLa cells against Stxs were followed in the presence of various concentrations of the peptide. Results were the average of at least four different samples.

Fig. 4 Representative computer-simulated binding mode between peptide (**WHWTW**) and Stx-1 B subunit. (a) Top view and (b) side view.

Tables

Table 1 Peptide sequence of phage clones selected (a) by biopanning to the Fab fragment of anti-Gb3 antibody, and (b) by biopanning with subtraction strategy.

	Clone	Sequence	Frequency
(a)	Gb3-1	WHWTWLSEYPPP	3/10
	Gb3-2	GGPIPTIPRPTL	2/10
	Gb3-3	WHWTWLSDYPPP	1/10
	Gb3-4	WQWTWLCEHPPP	1/10
	Gb3-5	WHRPPWWPSSSL	1/10
	Gb3-6	HELRSQIISTTS	1/10
	Gb3-7	FQWSWYTPPRPS	1/10
(b)	Gb3-S-1 (Gb3-1)	WHWTWLSEYPPP	3/10
	Gb3-S-2	EQWPWSLLPIHY	2/10
	Gb3-S-3	NALTSHTASPAP	1/10
	Gb3-S-4	HPPGHRLTLIPH	1/10
	Gb3-S-5	VPSLHQTTHTPTP	1/10
	Gb3-S-6	HPSCNATGIPH	1/10

Table 2 Kinetic constants of **AcWHWTWLSEYK** to Stxs estimated by SPR.

	$k_a(\text{M}^{-1} \text{s}^{-1})$	$k_d(\text{s}^{-1})$	$K_D (\text{M})$
Stx-1	1.2×10^4	1.6×10^{-5}	1.4×10^{-9}
Stx-2	4.8×10^3	1.1×10^{-5}	2.3×10^{-9}